

THE STRUCTURE OF CLEOMEOLIDE, AN UNUSUAL BICYCLIC DITERPENE FROM *CLEOME VISCOSA* L. (CAPPARACEAE)

BASIL A. BURKE, WILFRED R. CHAN^{5a} and VIDYA A. HONKAN

Department of Chemistry, The University of the West Indies, Mona, Kingston 7, Jamaica

and

JOHN F. BLOUNT and PERCY S. MANCHAND

Chemical Research Department, Hoffmann-La Roche Inc., Nutley, NJ 07110, U.S.A.

(Received in USA 6 November 1979)

Abstract—A bicyclic diterpene, cleomeolide (**1**), $C_{20}H_{30}O_3$, has been isolated from *Cleome viscosa* L. (Capparaceae), and its stereostructure established by chemical, spectral and X-ray crystallographic means. Crystals of **1** belong to space group $P2_1$, with $a = 10.329(2)\text{Å}$, $b = 12.468(3)\text{Å}$, $c = 7.356(2)\text{Å}$, $\beta = 109.98^\circ$, and $Z = 2$. The structure was solved by a multiple solution procedure and refined by full matrix least-squares to give $R = 0.041$ and $wR = 0.051$. Oxidation of **1** gave the ketone **4**, which on treatment with methanolic KOH led to a facile transannular reaction to give the lactone **6**.

The macrocyclic diterpenes cembrene¹ and casbene² may be regarded as the prototypes of a variety of natural products, many of which show pronounced biological activity.³ Examples of these compounds are the esters of phorbol (cocarcinogens),⁴ jatrophone (antileukemic),⁵ gnidimacrin (antileukemic),⁶ the *Pimelea* factors (irritants),⁷ asperdiol (antileukemic),⁸ taxol (antitumor, microtubule promotion),⁹ and the kansuines A and B (analgesic).¹⁰ In continuation of our research¹¹ in this area, we report the isolation and structural characterization of cleomeolide (**1**), a bicyclic diterpene with a skeleton related to that found in verticillol,¹² and also discuss some interesting reactions encountered during our structural investigation.

Cleomeolide (**1**), $C_{20}H_{30}O_3$, m.p. 216–217°, was isolated as colourless crystals from a benzene extract of the leaves and twigs of *Cleome viscosa* (syn. *C. icosandra*, *Polanisia viscosa*, Capparaceae),¹³ and had IR (KBr, cm^{-1}) absorptions indicative of OH (3435), exocyclic methylene (1640, 900), and α,β -unsaturated lactone (1683, 1628) groups. The single maximum in the UV spectrum at 214 nm (ϵ 4800) supported the presence of the latter functionality. The ¹H NMR spectrum of **1** displayed signals attributable to a secondary Me group (0.86, d, $J = 7$ Hz), a quaternary Me group (1.00, s), a Me group on a fully substituted C atom bearing an oxygen function (1.35, s), a methine proton of a secondary alcohol (4.53, br d, $J = 12$ Hz), a terminal methylene group (4.80 and 4.90, each a d, $J = 2$ Hz), and a vinyl proton (6.20, m). From the preceding spectral data and the molecular formula cleomeolide (**1**) is a bicarbocyclic diterpene. Additional structural information was obtained from the Fourier transform ¹³C NMR spectrum (Table 1), which confirmed the presence of three Me groups (δ at 15.8, 23.2, and 24.9 ppm), an exocyclic methylene

group (δ at 115.9 ppm), and a carbonyl of the ester type (δ at 170.0 ppm). Olefinic absorptions due to C-7, C-8, and C-15 occurred at 135.0, 139.2, and 150.0 ppm respectively. Further, there were absorptions due to two sp^3 carbons bearing oxygen (δ at 68.8 and δ at 84.7 ppm), seven methylene groups, and two tertiary carbons.

The secondary nature of the alcohol group in cleomeolide (**1**) was confirmed by acetylation with acetic anhydride in pyridine. This gave the crystalline acetate **2**, $C_{22}H_{32}O_4$, in which the proton absorption at δ 4.53 in **1** was shifted to δ 5.83 (br d, $J = 10$ Hz). Jones oxidation afforded the ketone **4**, $C_{20}H_{28}O_3$, ν_{max} 1710 cm^{-1} . The 1,2-relationship between the OH and lactone functionalities was established as follows. Treatment of **1** with 10% KOH in aqueous methanol followed by acidification with acetic acid and esterification of the product with diazomethane furnished the crystalline epoxide **5**, $C_{21}H_{32}O_3$, whose ¹H NMR showed absorption due to the epoxide proton at δ 3.13 (br d; $J = 10$ Hz). Formation of the epoxide **5** suggested that there was a *trans* relationship between the OH group and the lactone oxygen in cleomeolide. Acid treatment of **5** gave cleomeolide.

Two products were obtained when the ketone **4** was treated with 10% KOH at room temperature: an isomeric lactone, m.p. 190–192°, and the acid **7**. The former could be formulated as the tricyclic lactone **6** on the basis of its spectral properties. Thus, it had no maximum in its UV spectrum above 220 nm, while its IR spectrum showed absorption for OH (3540) and γ -lactone (1763) groups. Its ¹H NMR spectrum (Experimental) showed retention of the exocyclic methylene group, no signals for CHOH , but signals for a disubstituted olefin. The structure of **6** was strongly supported by its ¹³C NMR spectrum (Table 1) and was conclusively established by X-ray crystallographic analysis. A stereoscopic drawing of **6** is shown in Fig. 1, and the pertinent X-ray crystallographic data are given in Table 2. As can be seen from Fig. 1, **6** is a tricyclic compound, formed by a facile transannular reaction between C-3 and C-8 of **4**. The

^aPresent address: Department of Chemistry, The University of the West Indies, St. Augustine, Trinidad, West Indies.

Table 1. ^{13}C NMR data for (1) and (6)^a

Carbon atom	(1)	(6)
1	41.2 (d)	40.3 (d)
2	b	c
3	69.8 (d)	78.7 (s)
4	84.7 (s)	84.4 (s)
5	b	c
6	36.7 (t)	128.9 (d)
7	135.0 (d)	129.3 (d)
8	139.2 (s)	55.4 (s)
9	b	c
10	b	c
11	42.1 (s)	43.5 (s)
12	39.5 (d)	39.7 (d)
13	b	c
14	b	c
15	150.0 (s)	154.2 (s)
16	24.9 (q)	25.2 (s)
17	170.0 (s)	175.2 (s)
18	23.2 (q)	20.3 (q)
19	15.8 (q)	15.4 (q)
20	115.9 (t)	113.1 (t)

a. Determined in CDCl_3 at 25.2 MHz. Chemical shifts are in p.p.m. downfield from Me_4Si .

b. Unassigned CH_2 absorptions at 23.6, 24.9, 27.2, 29.2, 33.8, and 37.1 p.p.m.

c. Unassigned CH_2 absorptions at 24.8, 26.4, 27.4, 33.5, 33.8, and 36.8 p.p.m.

tricyclic system in **6** is analogous to that found in the taxane¹⁴ group of diterpenes and could be of some biosynthetic significance.

When the lactone **6** was boiled under reflux (1 hr) with 10% KOH in aqueous methanol, followed by acidification, the crystalline hydrocarbon **9**, $\text{C}_{19}\text{H}_{26}$, was produced. Formation of **9** most likely occurred via the diene **10** in a decarboxylation-elimination process. Indeed, careful neutralization of the reaction mixture led to the detection of an unstable substance whose UV spectrum (λ_{max} 277 nm) was consistent with **10**.

Although the preceding chemical and spectral evidence served to delineate the main structural

features of cleomeolide, it was considered insufficient to define unambiguously the stereostructure of this compound. Consequently, an X-ray structure determination of cleomeolide (**1**) was undertaken. A stereoscopic drawing of **1** as determined from the X-ray crystallographic analysis is displayed in Fig. 2. This Figure also represents the absolute stereochemistry as determined from application of Brewster's rule¹⁵ on the benzoate **3**. Details of the X-ray analysis are given in Table 2. Cleomeolide possesses a 12-membered carbocyclic ring *cis* fused to a cyclohexane ring. There is also a 7-membered α,β -unsaturated lactone in which the double bond is at the bridgehead position.¹⁶

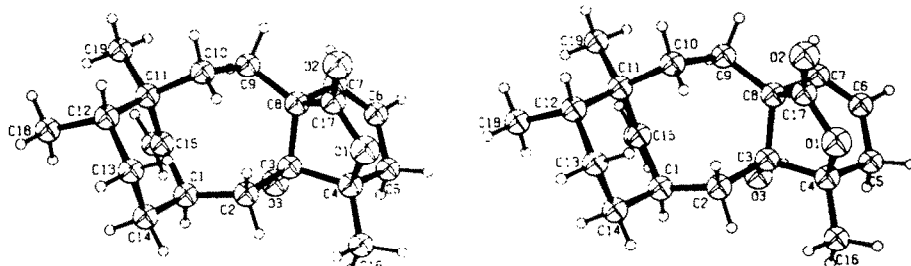


Fig. 1. A stereoscopic ORTEP drawing of lactone (**6**).

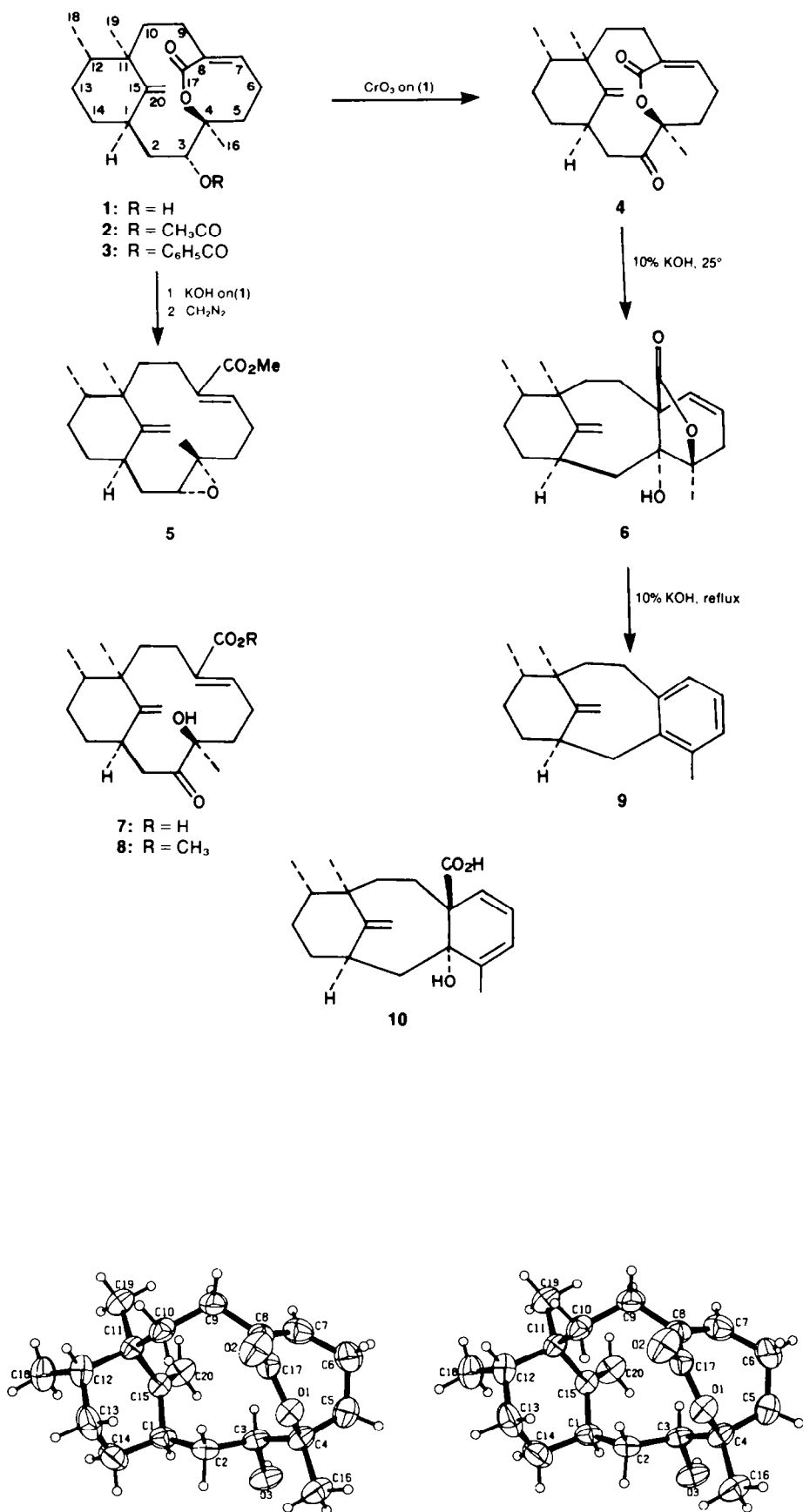
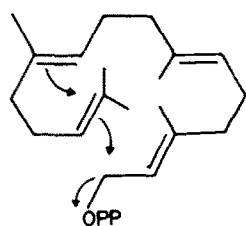


Table 2. Crystal data and details of X-ray crystallographic analyses for (1) and (6)

	(1)	(6)
Formula	C ₂₀ H ₃₀ O ₃	C ₂₀ H ₂₈ O ₃
Space group	P2 ₁	P2 ₁
a/Å	10.329 (2)	6.975 (1)
b/Å	12.468 (3)	11.700 (2)
c/Å	7.356 (2)	10.370 (1)
β/°	109.98 (2)	95.64 (1)
Z	2	2
D _{calcd.} /gm cm ⁻³	1.187	1.247
μ(Cu-Kα)/cm ⁻¹	6.2	6.6
Crystal size/mm	0.15x0.40x0.50	0.20x0.20x0.50
Max. θ/°	57	57
Total reflections	1899	1197
No. of obsd reflections	1859	1186
Final R	0.041	0.034
Final wR	0.051	0.049
Final difference map, largest peak/eÅ ⁻³	< ± 0.3	< ± 0.2

The genesis of the cleomeolide skeleton is readily rationalized in terms of head-to-tail cyclization of geranyl geranyl pyrophosphate followed by hydride shift, Me migration and proton loss, as indicated below.



EXPERIMENTAL

M.p.'s were determined on a Kofler hot-stage apparatus or in capillaries and are uncorrected. Unless otherwise indicated, UV and IR spectra were determined in EtOH and CHCl₃ respectively. NMR spectra were determined in CDCl₃ with Me₄Si as an internal standard. Chemical shifts are expressed in parts per million (δ) and coupling constants (J) in Hz (s = singlet, d = doublet, t = triplet, q = quartet, and m = multiplet). Mass spectra were recorded using a direct inlet system with an ionization energy of 70 eV; *m/e* values are given with relative intensities in parentheses.

Isolation of cleomeolide (1). Finely ground leaves and twigs of *Cleome viscosa* (3.8 kg), collected near the Mona campus, were extracted by cold percolation with benzene. The green gum (317 g), obtained by removal of the solvent under reduced pressure (below 50°), was chromatographed twice on alumina. Elution with benzene-EtOAc gave fractions which crystallized from MeOH to give cleomeolide (1) as prisms, m.p. 216–217°, [α]_D²⁵ + 120° (CHCl₃, *c* = 1.1); λ _{max} 214 nm (ε 4800); ν _{max} (KBr) 3485, 1683, 1642, 1628, 900 cm⁻¹; δ 0.86 (d, J = 7 Hz, CH₃-18), 1.00 (s, CH₃-19), 1.35 (s, CH₃-16), 4.53

(br d, J = 12 Hz, CH₃-3), 4.80 (d, J = 2 Hz, CH_A-20), 4.90 (d, J = 2 Hz, CH_B-20), 6.20 (m, CH₇-7); *m/e* 318 (M⁺, 3), 303 (M⁺-CH₃, 2), 300 (M⁺-H₂O, 9), 285 (4), 272 (8), 247 (7), 234 (8), 227 (6), 206 (13), 188 (25), 173 (12), 161 (12), 147 (16), 136 (25), 123 (100), 121 (60), 107 (35), 95 (30). (Found: C, 75.6; H, 9.3. C₂₀H₃₀O₃ requires: C, 75.4; H, 9.5%).

Cleomeolide acetate (2). This was prepared in the usual manner with Ac₂O and pyridine. It crystallized as needles from MeOH, m.p. 140–143°, [α]_D²⁵ + 96° (CHCl₃, *c* = 0.89), λ _{max} 216 nm (ε 4800); ν _{max} (CHCl₃) 1736, 1700, 900 cm⁻¹; δ 0.83 (d, J = 6.5 Hz, CH₃-18), 1.01 (s, CH₃-19), 1.33 (s, CH₃-16), 4.80 (d, J = 2 Hz, CH_A-20), 4.93 (d, J = 2 Hz, CH_B-20), 5.83 (br d, J = 10 Hz, CH₃-3), 6.13 (m, CH₇-7); *m/e* 360, 231. (Found: C, 73.5; H, 9.0. C₂₂H₃₂O₄ requires: C, 73.3; H, 8.9%).

Cleomeolide benzoate (3). This was prepared in the usual manner from benzoyl chloride and pyridine at room temp during 16 hr. Purification of the product by preparative scale tlc and crystallization from aqueous MeOH gave the benzoate (3), m.p. 157–159°, [α]_D²⁵ + 75° (CHCl₃, *c* = 1.00), ν _{max} 1710, 1630, 1600, 1580, 920 cm⁻¹; δ 0.83 (d, J = 7 Hz, CH₃-18), 1.03 (s, CH₃-19), 1.43 (s, CH₃-16), 4.97 (d, J = 2 Hz, CH_A-20), 5.03 (d, J = 2 Hz, CH_B-20), 6.17 (br d, J = 10 Hz, CH₃-3), 6.23 (m, CH₇-7), 7.50 (3 H, m, Ar-H), 8.07 (2 H, m, Ar-H). (Found: C, 76.55; H, 8.3. C₂₇H₃₄O₄ requires: C, 76.7; H, 8.1%).

Treatment of cleomeolide with alkali. A soln of cleomeolide (250 mg) in 3.4% methanolic KOH (29 mL) was refluxed for

2.5 hr. Most of the MeOH was removed *in vacuo* and, after dilution with H₂O (20 mL), the soln was acidified with HOAc. The product, recovered with EtOAc, was esterified with diazomethane. Evaporation of the solvent gave a gum (2 main components by tlc), which was separated by preparative-scale tlc to give *cleomelide 1* (41 mg) and the *methyl ester 5* (210 mg). The latter crystallized from aqueous MeOH as needles, m.p. 78–80°, λ_{\max} 218 nm (ϵ 5950); ν_{\max} (CHCl₃) 1706, 907 cm⁻¹; δ 0.83 (d, J = 7 Hz, CH₃-18), 0.90 (s, CH₃-19), 1.21 (s, CH₃-16), 3.13 (br d, J = 10 Hz, CH-3), 3.70 (s, CO₂CH₃), 4.91 (d, J = 2 Hz, CH_A-20), 5.08 (d, J = 2 Hz, CH_B-20), 5.16 (d of d, J = 4, 12 Hz, CH-7). (Found: C, 75.5; H, 9.6. C₂₁H₃₂O₃ requires: C, 75.9; H, 9.7%).

Oxidation of cleomeolide. Cleomeolide 1 (325 mg) in acetone (25 mL) was oxidized with Jones reagent at room temp. Removal of the solvent *in vacuo* followed by dilution with water (50 mL) gave a crystalline ppt of **4** (297 mg), needles from MeOH, m.p. 160–163°, λ_{\max} 218 nm (ϵ 5950); ν_{\max} (CHCl₃) 1710, 907 cm⁻¹; δ 0.86 (d, J = 7 Hz, CH₃-18), 1.01 (s, CH₃-19), 1.45 (s, CH₃-16), 4.81 (d, J = 2 Hz, CH_A-20), 5.00 (d, J = 2 Hz, CH_B-20), 5.98 (m, CH-7). (Found: C, 75.7; H, 8.8. C₂₀H₂₈O₃ requires: C, 75.9; H, 8.9%).

Base treatment of ketone (4). A soln of **4** (193 mg) in 10% KOH in MeOH (6.8 mL) was stirred at room temp for 4 hr. Most of the solvent was removed *in vacuo* and the residue diluted with H₂O (25 mL). The ppt was filtered off, washed well with H₂O, dried and recrystallized from MeOH to furnish the *tricyclic lactone (6)* as needles (75 mg), m.p. 190–192°, ν_{\max} (KBr) 3540, 1763, 1638, 887 cm⁻¹; δ 0.88 (d, J = 7 Hz, CH₃-18), 0.94 (s, CH₃-19), 1.34 (s, CH₃-16), 4.74 (d, J = 2 Hz, CH_A-20), 5.16 (d, J = 2 Hz, CH_B-20), 5.18 (d of t, J = 1, 9 Hz, CH-7), 5.86 (d of t, J = 4, 9 Hz, CH-6); MS: *m/e* 316 (M⁺, 5), 301 (5), 272 (10), 257 (7), 207 (25), 163 (45), 123 (56), 43 (100). (Found: C, 76.0; H, 8.9. C₂₀H₂₈O₃ requires: C, 75.9; H, 8.9%).

The filtrate from the preceding experiment was acidified with dil HCl and the resulting mixture was extracted with EtOAc (4 × 30 mL). The combined organic layers were washed with H₂O (2 × 20 mL) and evaporated to **7**, m.p. 98–100° (from aq. MeOH), which was characterized as its *methyl ester (8)* by treatment with CH₂N₂. It crystallized from aq. MeOH (needles), m.p. 126–127°, λ_{\max} 224 nm (ϵ 5200); ν_{\max} 3460, 1718, 1706 cm⁻¹; δ 0.83 (s, CH₃-18), 0.86 (d, J = 6.5 Hz, CH₃-19), 1.16 (s, CH₃-16), 3.80 (s, CO₂CH₃), 4.81 (d, J = 2 Hz, CH_A-20), 5.52 (d, J = 2 Hz, CH_B-20), 5.53 (m, CH-7). (Found: C, 72.35; H, 9.3. C₂₁H₃₂O₄ requires: C, 72.4; H, 9.3%).

The aromatic hydrocarbon (9). A soln of **6** (55 mg) and 10% KOH in MeOH was boiled under reflux for 1 hr. After removal of the solvent *in vacuo*, the mixture was diluted with H₂O (8 mL) and filtered. The filtrate was acidified with dil HCl and extracted with EtOAc (3 × 30 mL). The combined organic extracts were washed with H₂O (2 × 10 mL) and evaporated. The product **9** was then recrystallized from aqueous MeOH to give plates (20 mg), m.p. 87–89°, ν_{\max} 1631, 898 cm⁻¹; δ 0.86 (d, J = 7 Hz, CH₃-18), 0.91 (s, CH₃-19), 2.30 (s, CH₃-16), 4.13 (d, J = 2 Hz, CH_A-20), 4.45 (d, J = 2 Hz, CH_B-20), 6.93 (m, CH-5, CH-6, CH-7); *m/e* 254 (M⁺, 78), 249 (8), 226 (54), 212 (14), 197 (12), 184 (15), 171 (35), 156 (63), 143 (52), 131 (38), 123 (100), 118 (45), 107 (50), 91 (40), 81 (42) (M⁺ 254.2049; C₁₉H₂₆, requires: 254.2036).

X-Ray crystallography. The crystallographic data for **1** and **6** were collected on a fully-automated Hilger-Watts diffractometer (Cu-K α radiation, θ 2 θ scans, pulse height discriminations, no absorption corrections applied), and are summarized in Table 2. The structure and relative

stereochemistry of **1** and **6**, using crystals grown from MeOH, were solved by a multiple soln procedure¹⁷ and refined by full matrix least-squares. In the final refinement, anisotropic thermal parameters were used for non-H atoms and isotropic factors for the H atoms. H atoms were included in the structure factor calculation but their parameters were not refined. Listings of final atomic parameters, anisotropic thermal parameters, bond lengths, bond angles, torsion angles, and observed and calculated structure factors (22 p.p.) are submitted as supplementary material to be deposited in the Cambridge University Crystallographic Centre.¹⁸

REFERENCES

- W. G. Dauben, W. E. Thiessen and P. R. Resnick, *J. Am. Chem. Soc.* **84**, 2015 (1962); *J. Org. Chem.* **30**, 1693 (1965); W. G. Dauben, J. P. Hubbell, P. Oberhansli and W. E. Thiessen, *Ibid.* **44**, 669 (1979).
- D. R. Robinson and C. A. West, *Biochem.* **9**, 70 (1969); D. Sutton and C. A. West, *Phytochem.* **14**, 1921 (1975).
- A. J. Weinheimer, C. W. J. Chang and J. A. Matson, *Fortschr. Chem. Org. Naturst.* **36**, 285 (1979).
- J. D. White and P. S. Manchand, *Bio-organic Chemistry*, (Edited by E. E. van Tamelen), Vol. 2, p. 337. Academic Press, New York (1978). W. Adolf and E. Hecker, *Isr. J. Chem.* **16**, 75 (1977); E. Hecker, *Pure and Appl. Chem.* **49**, 1423 (1977); Y. Hirata, *Ibid.* **41**, 175 (1975).
- S. M. Kupchan, C. W. Sigel, M. J. Matz, C. J. Gilmore and R. F. Bryan, *J. Am. Chem. Soc.* **98**, 2295 (1976).
- S. M. Kupchan, Y. Shizuri, T. Murai, J. G. Sweeny, H. R. Haynes, M. S. Shen, J. C. Barrick, R. F. Bryan, D. Holm and K. K. Wu, *Ibid.* **98**, 5719 (1976).
- S. Zayed, W. Adolf, A. Hafez and E. Hecker, *Tetrahedron Letters* 3481 (1977).
- A. J. Weinheimer, J. A. Matson, D. Helm and M. Poling, *Ibid.* 1295 (1977).
- M. C. Wani, H. L. Taylor, M. E. Wall, P. Coggon and A. T. McPhail, *J. Am. Chem. Soc.* **93**, 2325 (1971).
- D. Uemura, Y. Hirata, Y. P. Chen and H. Y. Hsu, *Tetrahedron Letters* 1697 (1975); D. Uemura, C. Katayama, E. Uno, K. Sasaki, Y. Hirata, P. Chen and H. Y. Hsu, *Ibid.* 1703 (1975); D. Uemura and Y. Hirata, *Ibid.* 1701 (1975).
- P. S. Manchand and J. F. Blount, *J. Org. Chem.* **42**, 3824 (1977); W. R. Chan, E. C. Prince, P. S. Manchand, J. P. Springer and J. Clardy, *J. Am. Chem. Soc.* **97**, 4437 (1975); B. A. Burke, W. R. Chan, K. O. Pascoe, J. F. Blount and P. S. Manchand, *Tetrahedron Letters* 3345 (1979).
- B. Karlsson, A.-M. Pilotti, A.-M. Söderholm, T. Norin, S. Sundin and M. Sumimoto, *Tetrahedron* **34**, 2349 (1978).
- C. D. Adams, *Flowering Plants of Jamaica*, p. 303. University of the West Indies, Mona, Jamaica (1972).
- K. Nakanishi, T. Goto, S. Ito, S. Natori and S. Nozoe, *Natural Products Chemistry*, Vol. 1, p. 281. Academic Press, New York (1974).
- J. Brewster, *Tetrahedron* **13**, 106 (1961).
- During the preparation of our manuscript a paper appeared in which the X-ray structure of **1** was disclosed (S. B. Mahato, B. C. Pal, T. Kawasaki, K. Miyahara, O. Tanaka and K. Yamasaki, *J. Am. Chem. Soc.* **101**, 4720 (1979)).
- G. Germain, P. Main and M. M. Woolfson, *Acta Crystallogr. Sect. A*, **27**, 368 (1971).
- See *Tetrahedron* **35**, 448 (1979).